



Optimizing Human Pluripotent Stem Cell (hiPSC) Production Process Using Vertical-Wheel Bioreactors

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Vertical-Wheel Stirred Suspension Bioreactors

Growth Platforms

Bioreactor

X Variability between flasks

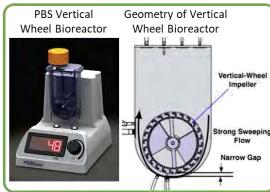
Static

- More difficult to regulate culture environment
- X Labour intensive
- X Batch or fed batch mode only

- Well-mixed vessel
- Less labour intensive
- Easier to monitor and control process parameters
- Can operate in batch, perfusion mode, etc.
- Scalable

Vertical-Wheel Reactors

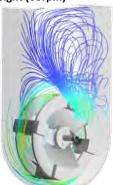




Velocity though the Reactor Height (60rpm)





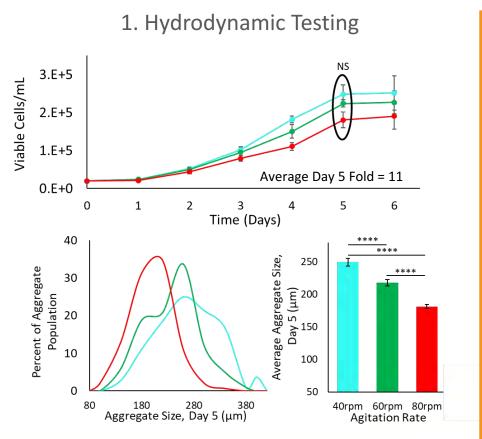




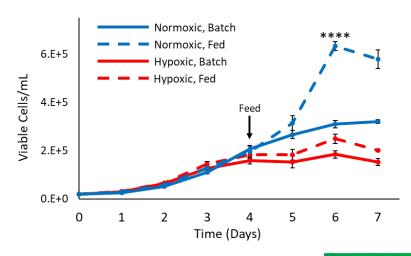
Growth of hiPSCs in PBS-0.1

Inoculation of hiPSC Aggregates Pre-Formed in Static Culture

Early Studies by our Team using Spinners and Stirred-Tank Bioreactors: Unsuccessful cell growth and poor aggregate distributions when cells were inoculated as a single cell suspension. For this reason, the team developed a passaging method involving a pre-formation of hiPSCs in static culture prior to inoculating cells into a bioreactor.



2. Oxygen and Nutrient Testing

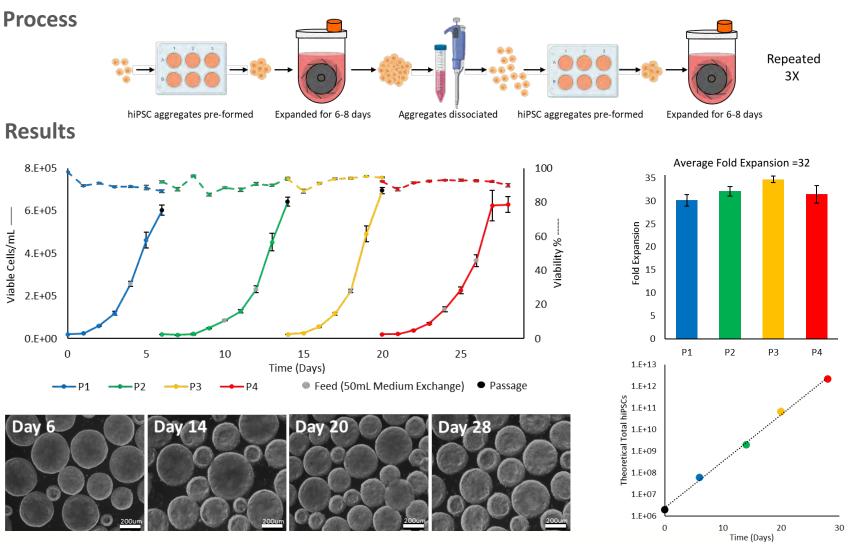


Reference	Bioreactor Type	Fold/Days
Zweiggerdt et al. (2011)	Horizontal Blade	3-6 fold / 4-7 days
Abbasalizadeh et al. (2012)	Horizontal Blade	8 fold / 7-10 days
Haraguchi et al. (2015)	Horizontal Blade	10 fold / 12 days
Badenes et al. (2016)	Horizontal Blade	3.5 fold / 10 days
This Study	Vertical Wheel	32 fold / 6 days



Successful Serial Passage of hiPSCs in PBS 0.1L

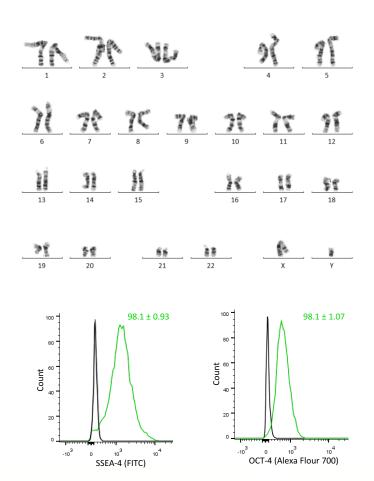
Achieved a Total of >1E6-Fold Expansion (n=4)

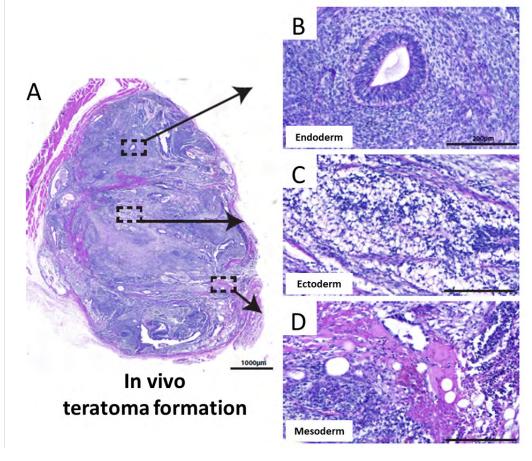




Serial Passaged hiPSC are of High Quality

After 28 days of VW bioreactor culture, hiPSCs maintain normal karyotype and characteristic pluripotent stem cell phenotype and function

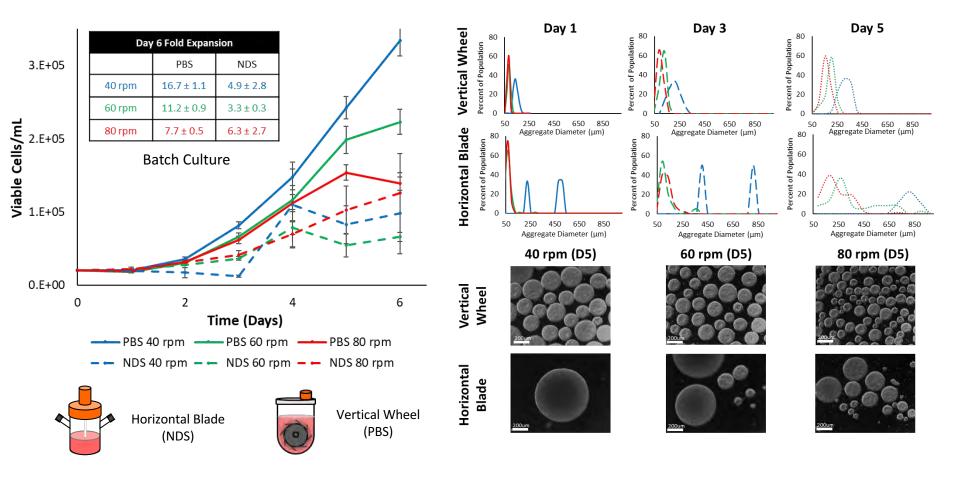






Single Cell hiPSC Inoculation in PBS-0.1 Vessels vs Spinner (NDS)

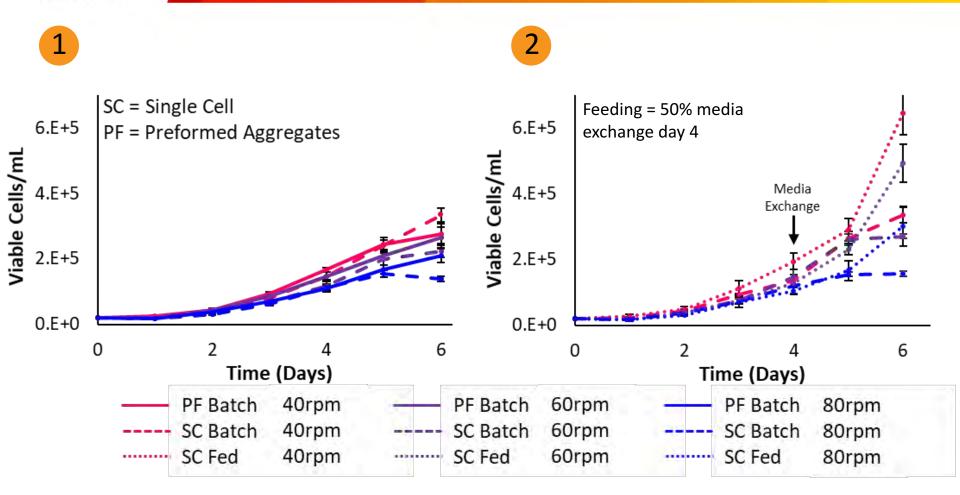
Single cell inoculation led to successful cell growth in vertical-wheel bioreactors but not in traditional horizontal-blade bioreactors



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1. Single Cell vs Pre-formed Aggregate Inoculation in PBS 0.12. Single Cell Batch vs Fed-Batch in PBS 0.1

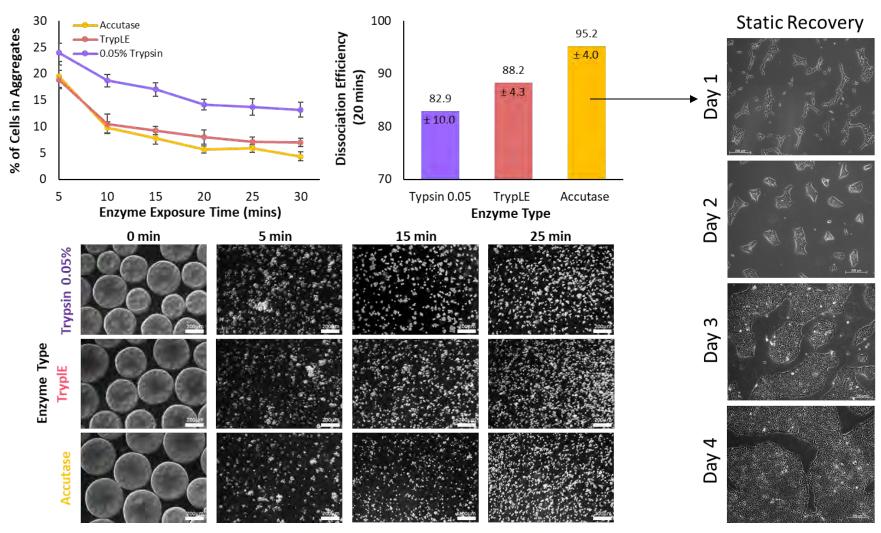


1. Single cell and pre-formed aggregate inoculation resulted in similar growth in the PBS-0.1 at all agitation rates

2. Fed-batch culture resulted in approximately 2X higher fold expansion at all agitation rates

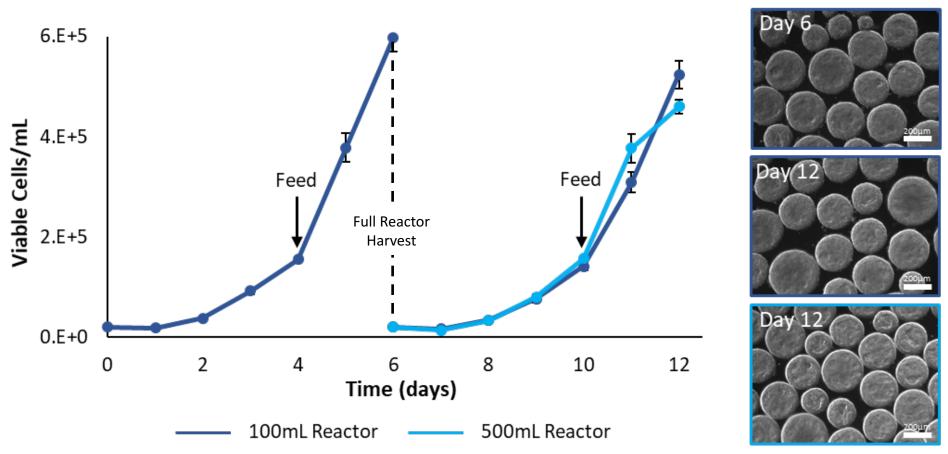


Test of Proteolytic Enzymes and Dissociation Time with Agitation





Combined Process (Single Cell & Bioreactor Harvest) Serial Passage in PBS-0.1 and PBS-0.5



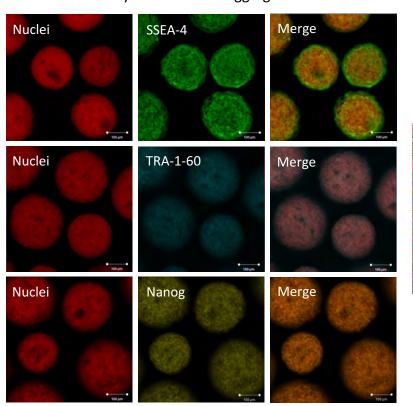
hiPSC cell growth remains high after bioreactor harvest and re-seeding of single cells into PBS 0.1 and 0.5



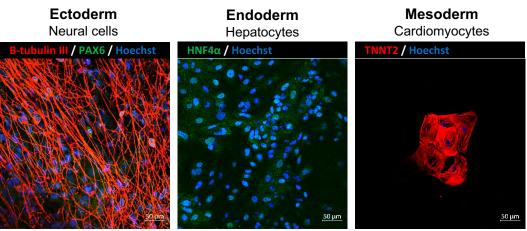
Combined Process (Single Cell & Bioreactor Harvest) Serial Passage in PBS-0.1 and PBS-0.5

hiPSC maintain high quality pluripotent characteristics following optimized serial passage protocol with single cell inoculation and full bioreactor harvesting

Pluripotency Staining Day 12 Bioreactor Aggregates



Tri-lineage Differentiation



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1. Optimizing bioprocess variables (agitation, oxygen, and nutrients) in the PBS-0.1L reactor resulted in hiPSC expansion far greater than previously reported

2. The expansion process is highly reproducible with the potential to generate over 1E12 high quality PSC (starting from 2E6) over 4 serial passages

3. Bioprocess variables within the PBS reactor can be altered to overcome bioprocess bottlenecks in manufacturing hiPSCs (namely single cell seeding and full bioreactor harvests)

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